PUBLIC NOTICE

Initiation of Risk Assessments for Chemicals in Drinking Water [09/20/07]

A. Requirements

The Calderon-Sher California Safe Drinking Water Act of 1996 requires the Office of Environmental Health Hazard Assessment (OEHHA) to post notices on its Web site of water contaminants for which it is initiating development of public health goals (PHGs) for the chemicals in drinking water. The law also describes the intent and general context of the PHGs. PHGs are concentrations of chemicals in drinking water that are not anticipated to produce adverse health effects following long-term exposures. OEHHA is required to consider potential adverse effects on members of subgroups that comprise a meaningful proportion of the population, including but not limited to infants, children, pregnant women, the elderly, and individuals with a history of serious illness. The public health goals are non-regulatory in nature but are to be used as the health basis to update the state's primary drinking water standards (maximum contaminant levels, or MCLs) established by the California Department of Public Health (DPH) for chemicals subject to regulation.

The act requires PHGs to be developed for the approximately 88 chemicals for which state or federal MCLs are provided, and review and update the risk assessments that form the basis for the PHGs as appropriate at least every five years. Other chemicals may be added to the list by legislative or interdepartmental request. Opportunities for public comment and peer review are provided.

B. Implementation

OEHHA has published 80 PHGs as of September 2007. Two MCLs, for gross alpha and gross beta radionuclides, represent screening levels for contaminants rather than specific regulatory standards; for these, OEHHA has provided risk assessments and guidance memoranda. The technical support documents for these chemicals are posted on the OEHHA Web site at www.oehha.ca.gov.

In addition, PHG re-evaluations have been completed for eight chemicals. For two of these chemicals (cadmium and glyphosate), a complete new PHG document was prepared and published. For the six other chemicals (chlordane, 1,2-dichloroethane, 1,3-dichloropropene, inorganic mercury, lindane, and thallium), OEHHA concluded that no new information was available on these chemicals that would require significant changes to the PHG document. Memoranda to this effect are available at http://www.oehha.ca.gov/water/reports/index.html. The re-reviews of several other chemicals that were announced previously are in progress. Draft PHG documents for the remaining chemicals with existing MCLs or requests for development of a PHG are in preparation.

Draft PHG documents on three chemicals (chlorite, PCBs, and TCDD) have been posted on the OEHHA Web site for public comment, along with drafts of PHG updates for copper and 2,4-D. A 45-day public comment period is provided after posting of the initial drafts, along with a public workshop to elicit comments and discussion. After

revision with consideration of comments received, a further 30-day public comment period will follow. The final revision will include responses to major comments.

Evaluation is now being initiated for several other chemicals for which PHGs were developed earlier (see Section D), which are being re-reviewed as part of the ongoing PHG update process. Information relevant to the development of PHGs is requested on each of these chemicals.

C. PHGs to be released for public review:

Draft documents for the following "new" chemicals (those with no existing PHG) are in progress and are planned for release for initial public review and comment when they are completed:

- Bromate
- Haloacetic acids
- Hexavalent chromium
- Molinate
- Selenium
- Styrene
- 1,2,3-Trichloropropane
- Trihalomethanes

Toxicity reviews are in progress for the following additional chemicals, for which initiation of review was previously announced:

- Alachlor
- Atrazine and simazine
- Chlorite
- Copper
- 2,4-D (Dichlorophenoxyacetic acid)
- Fluoride
- Lead
- Nitrate/nitrite
- PCBs (Polychlorinated biphenyls)
- Trichloroethylene

D. Initiation of risk assessments

Risk assessment is being initiated for the following list of chemicals:

- Antimony
- Bentazon
- Benzo(a)pyrene
- Cyanide
- Dalapon
- Dibromochloropropane (DBCP)
- 1,2-Dichlorobenzene

- 1,4-Dichlorobenzene
- 1,1-Dichloroethylene
- 1,2-Dichloropropane
- Dinoseb
- Endothall
- Endrin
- Hexachlorocyclopentadiene
- Methoxychlor
- Oxamyl
- Pentachlorophenol
- Picloram
- 1,2,4-Trichlorobenzene
- Trichlorofluoromethane (Freon 11)
- Trichlorotrifluoroethane (Freon 113)

These risk assessments are updates of assessments prepared in the first two years of our program (1997 and 1999). The chemicals had earlier been prioritized for review on the basis of availability of new data and significance as drinking water contaminants, so these chemicals on the final list were considered of lowest priority. Little new data are available for most of them. A brief description of the chemicals is provided below. This announcement solicits the submission of other pertinent information on the contaminants that could assist our office in preparing or updating the risk assessment and deriving a revised PHG.

Information submitted to OEHHA in response to this request should not be proprietary in nature, because all information submitted is a matter of public record. Information should be submitted by November 9, 2007 to:

Thomas Parker
PHG Project
Pesticide and Environmental Toxicology Branch
Office of Environmental Health Hazard Assessment
P.O. Box 4010
Sacramento, California 95812-4010

All data submitted will be considered in the development of the PHG for these chemicals. If substantive revisions to the original PHG documents are required, the draft documents will be available for discussion in a public workshop and public comment will be solicited as described above in Section B. The final risk assessments will be utilized by the DPH for potential revisions to the MCLs for the chemicals in drinking water, as described in more detail on their Web site at

http://www.dhs.ca.gov/ps/ddwem/chemicals/chemindex.htm.

E. Descriptions of chemicals or substances for initiation of review:

Antimony

In 1997, OEHHA developed a PHG for antimony and its compounds of 20 parts per billion (ppb) in drinking water. Antimony is an element found throughout the earth, but present in small quantities in ores. Since antiquity, antimony has been used for many purposes: as a cosmetic, drug, in utensils and currently as a fire retardant and a component in plastics. Antimony is usually available as a compound, and various antimony compounds behave differently in the environment and also have different effects upon humans. Antimony has been reported as exceeding the MCL in California public drinking water supplies eleven times between June 2002 and June 2006.

Ingestion of one antimony compound will produce severe stomach upset resulting in vomiting. Antimony fumes and dusts inhaled by workers are associated with the development of benign tumors of the lungs, dermatitis and, less commonly, effects on the heart and kidneys. Laboratory animals exposed to antimony by inhalation or ingestion exhibit effects similar to those noted in humans. However, antimony and its compounds are not considered to be carcinogenic. The original PHG for antimony was calculated based on minor clinical signs and a slight decrease in longevity noted in a chronic oral study conducted in rats. Other information used to develop the PHG included estimates of human exposure to antimony derived from measurements taken of levels of antimony in air, food and water. The studies presented below represent a selection of the available data.

Selected References

Andrewes P, Kitchin KT, Wallace K (2004). Plasmid DNA damage caused by stibine and trimethylstibine. Toxicol Appl Pharmacol 194:41-48.

Cavallo D, Iavicoli I, Setini A, Marinaccio A, Perniconi B, Carelli G, *et al.* (2002). Genotoxic risk and oxidative DNA damage in workers exposed to antimony trioxide. Environ Mol Mutagen 40:184-189.

De Boeck M, Kirsch-Volders M, Lison D (2003). Cobalt and antimony: Genotoxicity and carcinogenicity. Mutat Res 533:135-152.

Hext PM, Pinto PJ, Rimmel BA (1999). Subchronic feeding study of antimony trioxide in rats. J Appl Toxicol 19:205-209.

Khalil EA, Ahmed AE, Musa AM, Hussein MH (2006). Antimony-induced cerebellar ataxia. Saudi Med J 27:90-92.

Kirkland D, Whitwell J, Deyo J, Serex T (2007). Failure of antimony trioxide to induce micronuclei or chromosomal aberrations in rat bone-marrow after sub-chronic oral dosing. Mutat Res 627:119-128.

Poon R, Chu I, Lecavalier P, Valli VE, Foster W, Gupta S, *et al.* (1998). Effects of antimony on rats following 90-day exposure via drinking water. Food Chem Toxicol 36:21-35.

Schaumloffel N, Gebel T (1998). Heterogeneity of the DNA damage provoked by antimony and arsenic. Mutagenesis 13:281-286.

Tarabar AF, Khan Y, Nelson LS, Hoffman RS (2004). Antimony toxicity from the use of tartar emetic for the treatment of alcohol abuse. Vet Hum Toxicol 46:331-333.

Bentazon

A PHG of 200 ppb was developed in 1999 for the herbicide bentazon based on a chronic exposure study in dogs. Bentazon is a benzothiadiazinone contact herbicide that acts as an inhibitor of photosynthetic electron transfer in plants. It has a low binding affinity to soil and thus has been found in ground and surface water in California. The detected levels ranged from 0.01 to $20.0~\mu g/L$ in 64 out of the 200 wells sampled. The herbicide degrades quickly in plants and soil (3-21 day half life); therefore any contamination of ground or surface water is likely to result from improper agricultural practices.

In the chronic dog study (Allen *et al.*, 1989), a NOAEL was established at 100 ppm in the food (approximately 3.2 mg/kg-day), and the LOAEL was 400 ppm (approximately 13.1 mg/kg-day). Adverse effects reported were emaciation, loose and/or bloody stools, pale mucous membranes, hematological changes suggestive of anemia, intestinal inflammation, and congestion of the small intestine and spleen. The anemia appeared to be due to blood loss from the gastrointestinal tract. This NOAEL was used to calculate the PHG.

Bentazon was classified by U.S. EPA as "Group E," meaning there was no evidence of carcinogenicity. However, two studies warrant a reexamination of the toxicity of bentazon: one, initiated in response to a perceived cancer cluster in a bentazon manufacturing plant, showed a small but statistically significant increase in prostate and stomach cancers, while the second reported mutagenicity in a fruit fly wing spot test (Ott *et al.*, 2006; Kaya *et al.*, 2004). Information on other applicable and relevant studies is requested.

References:

Allen TR, Frei T, Luethemeier H, Vogel O, *et al.* (1989). 52-Week oral toxicity (feeding) study with Bentazon technical in the dog. RCC Research and Consulting Co. AG; RCC project No. 067746. Available also as U.S. EPA MIRD No. 41054901.

Garagna S, Vasco C, Merico V, Esposito A, Zuccotti M, Redi CA (2005). Effects of a low dose of bentazon on spermatogenesis of mice exposed during foetal, postnatal and adult life. Toxicology 212(2-3):165-74.

Kaya B, Marcos R, Yanikoğlu A, Creus A (2004). Evaluation of the genotoxicity of four herbicides in the wing spot test of Drosophila melanogaster using two different strains. Mutat Res 557(1):53-62.

Lebailly P, Vigreux C, Lechevrel C, Ledemeney D, *et al.* (1998). DNA damage in mononuclear leukocytes of farmers measured using the alkaline comet assay: Modifications of DNA damage levels after a one-day field spraying period with selected pesticides. Canc Epid Biomark Prev 7(10):929-940.

Lin TJ, Hung DZ, Hu WH, Yang DY, Wu TC (1999). Acute basagran poisoning mimicking neuroleptic malignant syndrome. Hum Exp Toxicol 18(8):493-4.

Ott MG, Poche SL, Klees JE, Conner PR (2006). Investigation of cancer occurrences associated with an herbicide manufacturing facility. J La State Med Soc 158(5):239-48.

Pistl J, Kovalkovicová N, Holovská V, Legáth J, Mikula I (2003). Determination of the immunotoxic potential of pesticides on functional activity of sheep leukocytes in vitro. Toxicology 188:73–81.

Turcant A, Harry P, Cailleux A, Puech M, Bruhat C, Vicq N, Le Bouil A, Allain P (2003). Fatal acute poisoning by bentazon. J Anal Toxicol 27(2):113-7.

Benzo(a)pyrene

The PHG of 4 parts per trillion (ppt; $0.004 \mu g/L$) for benzo(a)pyrene (B(a)P) was published in December, 1997. B(a)P is one of many polycyclic aromatic hydrocarbons that form when organic matter burns incompletely. As such, B(a)P is a ubiquitous environmental contaminant that can be detected in air, water and soil and food. Exposure may occur due to ingestion, inhalation or dermal contact with contaminated air, water, soil or food.

Adverse effects on the hematological, gastrointestinal and immunological systems and effects on reproduction and development have been attributed to exposure to benzo(a)pyrene. Exposure of experimental animals to B(a)P has resulted in statistically significant increases in skin, lung, and forestomach tumors. A cancer potency factor based on an increase in forestomach tumors in mice was employed to develop the PHG for benzo(a)pyrene.

There are many new genotoxicity and mechanism studies of B(a)P, because B(a)P is considered a prototypical genotoxic carcinogen. Also, since the publication of the PHG for B(a)P, two animal studies have been conducted that provide cancer dose-response data for lifetime oral exposures to B(a)P.

Selected References

Aoki Y, Hashimoto AH, Amanuma K, Matsumoto M, *et al.* (2007). Enhanced spontaneous and benzo(a)pyrene-induced mutations in the lung of Nrf2-deficient gpt delta mice. Cancer Res 67(12):5643-8.

Brown LA, Khousbouei H, Goodwin JS, Irvin-Wilson CV, Ramesh A, Sheng L, McCallister MM, Jiang GC, Aschner M, Hood DB (2007). Down-regulation of early ionotrophic glutamate receptor subunit developmental expression as a mechanism for observed plasticity deficits following gestational exposure to benzo(a)pyrene. Neurotoxicology May 21 [Epub ahead of print].

Culp SJ, Gaylor DW, Sheldon WG, Goldstein LS, Ireland FA (1998). A comparison of the tumors induced by coal tar and benzo[a]pyrene in a 2-year bioassay. Carcinogenesis 19:117-124.

Hu Y, Bai Z, Zhang L, Wang X, Zhang L, Yu Q, Zhu T (2007). Health risk assessment for traffic policemen exposed to polycyclic aromatic hydrocarbons (PAHs) in Tianjin, China. Sci Total Environ 382(2-3):240-50.

Kassie F, Anderson LB, Scherber R, Yu N, Lahti D, Upadhyaya P, Hecht SS (2007). Indole-3-carbinol inhibits 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone plus benzo(a)pyrene-induced lung tumorigenesis in A/J mice and modulates carcinogen-induced alterations in protein levels. Cancer Res 67(13):6502-11.

Kommaddi RP, Turman CM, Moorthy B, Wang L, Strobel HW, Ravindranath V (2007). An alternatively spliced cytochrome P4501A1 in human brain fails to bioactivate polycyclic aromatic hydrocarbons to DNA-reactive metabolites. J Neurochem 102(3):867-77.

Konstandi M, Harkitis P, Thermos K, Ogren SO, Johnson EO, Tzimas P, Marselos M (2007). Modification of inherent and drug-induced dopaminergic activity after exposure to benzo(alpha)pyrene. Neurotoxicology May 6 [Epub ahead of print].

Kroese ED, Muller JJA, Mohn GR, Dortant PM, Wester PW (2001). Tumorigenic effects in Wistar rats orally administered benzo[a]pyrene for two years (gavage studies). Implications for human cancer risks associated with oral exposure to polycyclic aromatic hydrocarbons. Report 658603 010. National Institute of Public Health and the Environment. Bilthoven, The Netherlands.

Kummer V, Maskova J, Zraly Z, Matiasovic J, Faldyna M (2007). Effect of postnatal exposure to benzo[a]pyrene on the uterus of immature rats. Exp Toxicol Pathol Jun 19 [Epub ahead of print].

Kushman ME, Kabler SL, Ahmad S, Doehmer J, Morrow CS, Townsend AJ (2007). Protective efficacy of hGSTM1-1 against B[a]P and (+)- or (-)-B[a]P-7,8-dihydrodiol cytotoxicity, mutagenicity, and macromolecular adducts in V79 cells co-expressing hCYP1A1. Toxicol Sci May 24 [Epub ahead of print].

Lee BM, Shim GA (2007). Dietary exposure estimation of benzo[a]pyrene and cancer risk assessment. J Toxicol Environ Health A 70(15-16):1391-4.

Li D, Wang LE, Chang P, El-Naggar AK, Sturgis EM, Wei Q (2007). In vitro benzo[a]pyrene diol epoxide-induced DNA adducts and risk of squamous cell carcinoma of head and neck. Cancer Res 67(12):5628-34.

Marczynski B, Raulf-Heimsoth M, Spickenheuer A, Mensing T, Welge P, Forster K, Angerer J, Pesch B, Bramer R, Kafferlein HU, Breuer D, Hahn JU, Bruning T (2007). Ambient and biological monitoring of exposure and genotoxic effects in mastic asphalt workers exposed to fumes of bitumen. J Occup Environ Hyg 4 Suppl 1:127-36.

Rundle A, Madsen A, Orjuela M, Mooney L, Tang D, Kim M, Perera F (2007). The association between benzo[a]pyrene-DNA adducts and body mass index, calorie intake and physical activity. Biomarkers 12(2):123-32.

Sanyal MK, Li YL (2007). Deleterious effects of polynuclear aromatic hydrocarbon on blood vascular system of the rat fetus. Birth Defects Res B Dev Reprod Toxicol July 5 [Epub ahead of print].

Yao B, Fu J, Hu E, Qi Y, Zhou Z (2007). The Cdc25A is involved in S-phase checkpoint induced by benzo(a)pyrene. Toxicology 237(1-3):210-7.

Cyanide

The Public Health Goal (PHG) of 150 ppb for cyanide was published by OEHHA in December 1997. Cyanide is a naturally occurring chemical that is found in many commonly consumed foods such as almonds, lima beans, cassava, and mustard. Cyanide may also be present at low concentrations in source water as well as finished municipal water supplies. It can be derived from various industrial operations or produced by burning nitrogen-containing materials. High-doses of cyanide inhibit cellular enzymes resulting in hypoxia. Central nervous system (CNS) effects of prolonged or high-level exposure can include demyelinating lesions of the brain and Parkinsonian-like symptoms. The heart is also sensitive to cyanide induced hypoxia. However, cyanide intoxication from the consumption of drinking water is extremely uncommon.

In the survey of the literature, many additional studies relating to the effects of cyanide have been found since the publication of the PHG in 1997. The new toxicity information appears likely to have significant impact on the existing toxicology and risk assessment sections of the PHG document. The studies presented below represent a selection of the available data.

Selected References

Alarie Y (2002). Toxicity of fire smoke. Crit Rev Toxicol 32(4):259-89.

Beasley DM, Glass WI (1998). Cyanide poisoning: pathophysiology and treatment recommendations. Occup Med (Lond) 48(7):427-31.

Bhattacharya R, Vijayaraghavan R (2002). Promising role of alpha-ketoglutarate in protecting against the lethal effects of cyanide. Hum Exp Toxicol 21(6):297-303.

Borron SW, Stonerook M, Reid F (2006). Efficacy of hydroxocobalamin for the treatment of acute cyanide poisoning in adult beagle dogs. Clin Toxicol (Phila) 44 Suppl 1:5-15.

Chaturvedi AK, Sanders DC, Endecott BR, Ritter RM (1995). Exposures to carbon monoxide, hydrogen cyanide and their mixtures: interrelationship between gas exposure concentration, time to incapacitation, carboxyhemoglobin and blood cyanide in rats. J Appl Toxicol 15(5):357-63.

Eckstein M, Maniscalco PM (2006). Focus on smoke inhalation--the most common cause of acute cyanide poisoning. Prehospital Disaster Med 21(2 Suppl 2):s49-55.

Eisler R, Wiemeyer SN (2004). Cyanide hazards to plants and animals from gold mining and related water issues. Rev Environ Contam Toxicol 183:21-54.

Gracia R, Shepherd G (2004). Cyanide poisoning and its treatment. Pharmacotherapy 24(10):1358-65.

Nelson L (2006). Acute cyanide toxicity: mechanisms and manifestations. J Emerg Nurs 32(4 Suppl):S8-11.

Ozolu RI, Okolie NP, Ebeigbe AB, Karikari N (2007). Effects of sub-chronic oral cyanide on endothelial function in rabbit aortic rings. Hum Exp Toxicol 26(2):105-10.

Rachinger J, Fellner FA, Stieglbauer K, Trenkler J (2002). MR changes after acute cyanide intoxication. AJNR Am J Neuroradiol 23(8):1398-401.

Soto-Blanco B, Gorniak SL (2004). Prenatal toxicity of cyanide in goats--a model for teratological studies in ruminants. Theriogenology 62(6):1012-26.

Sousa AB, Soto-Blanco B, Guerra JL, Kimura ET, Gorniak SL (2002). Does prolonged oral exposure to cyanide promote hepatotoxicity and nephrotoxicity? Toxicology 174(2):87-95.

Tulsawani R, Bhattacharya R (2006). Effect of alpha-ketoglutarate on cyanide-induced biochemical alterations in rat brain and liver. Biomed Environ Sci 19(1):61-6.

Tulsawani R, Kumar D, Bhattacharya R (2007). Effect of pre-treatment of alphaketoglutarate on cyanide-induced toxicity and alterations in various physiological variables in rodents. Biomed Environ Sci 20(1):56-63.

Tulsawani RK, Debnath M, Pant SC, Kumar O, Prakash AO, Vijayaraghavan R, Bhattacharya R (2005). Effect of sub-acute oral cyanide administration in rats: protective efficacy of alpha-ketoglutarate and sodium thiosulfate. Chem Biol Interact 156(1):1-12.

Dibromochloropropane

A PHG for 1,2-dibromo-3-chloropropane (DBCP) of 1.7 ppt was developed by OEHHA in 1999, based on carcinogenicity in experimental animals in multiple toxicity studies. DBCP was used as a soil fumigant for multiple crops until suspension of its use in 1977 and final cancellation in 1985. DBCP migrated readily to groundwater and is very persistent in the aquifer; it has contaminated groundwater over a wide area in California's Central Valley. Although the most heavily contaminated sources are not used for drinking water, they may still be used for irrigation. Because of the volatility of DBCP, significant residues in crops are not expected, but some evaporation to air will occur.

DBCP caused forestomach cancers in rats and mice after gavage administration, as well as mammary gland tumors in female rats. It also caused oral and nasal cavity tumors after inhalation exposures in rats and mice, and lung tumors in mice. It is mutagenic and clastogenic. DBCP inhibits sperm production in humans and animals, and has been associated with human infertility after occupational exposures. The few studies of DBCP effects since publication of the PHG in 1999 may provide significant new perspective on mechanism of action of DBCP.

Selected References

Allen RH, Gottlieb M, Clute E, Pongsiri MJ, Sherman J, Obrams GI (1997). Breast cancer and pesticides in Hawaii: the need for further study. Environ Health Perspect 105 Suppl 3:679-83.

Clark HA, Snedeker SM (2005). Critical evaluation of the cancer risk of dibromochloropropane (DBCP). J Environ Sci Health C Environ Carcinog Ecotoxicol Rev 23(2):215-60.

Hofmann J, Guardado J, Keifer M, Wesseling C (2006). Mortality among a cohort of banana plantation workers in Costa Rica. Int J Occup Environ Health 12(4):321-8.

Holme JA, Bjorge C, Trbojevic M, Olsen AK, Brunborg G, Soderlund EJ, Bjoras M, Seeberg E, Scholz T, Dybing E, Wiger R (1998). Effects of chemical-induced DNA damage on male germ cells. Arch Toxicol Suppl 20:151-60.

IARC (1999). 1,2-Dibromo-3-chloropropane. IARC Monogr Eval Carcinog Risks Hum 71 Pt 2:479-500.

Lamb JC, Reel J, Tyl R, Lawton AD (1997). Dibromochloropropane. Summary of NTP reproductive toxicity study in mice. Environ Health Perspect 105 Suppl 1:299-300.

Meistrich ML, Wilson G, Porter KL, Huhtaniemi I, Shetty G, Shuttlesworth GA (2003). Restoration of spermatogenesis in dibromochloropropane (DBCP)-treated rats by hormone suppression. Toxicol Sci 76(2):418-26.

Meistrich ML, Wilson G, Shuttlesworth GA, Porter KL (2003). Dibromochloropropane inhibits spermatogonial development in rats. Reprod Toxicol 17(3):263-71.

National Toxicology Program (2005). 1,2-Dibromo-3-chloropropane. 11th Report on Carcinogens. Accessed at: http://ntp.niehs.nih.gov/ntp/roc/eleventh/profiles/s058dibr.pdf.

Ryu JC, Kim YJ, Chai YG (2002). Mutation spectrum of 1,2-dibromo-3-chloropropane, an endocrine disruptor, in the lacI transgenic Big Blue Rat2 fibroblast cell line. Mutagenesis 17(4):301-7.

Sasaki YF, Saga A, Akasaka M, Ishibashi S, Yoshida K, Su YQ, Matsusaka N, Tsuda S (1998). Detection of in vivo genotoxicity of haloalkanes and haloalkenes carcinogenic to rodents by the alkaline single cell gel electrophoresis (comet) assay in multiple mouse organs. Mutat Res 419(1-3):13-20.

Spira A, Multigner L (1998). The effect of industrial and agricultural pollution on human spermatogenesis. Hum Reprod 13(8):2041-2.

Sussman NB, Mazumdar S, Mattison DR (1999). Modeling adverse environmental impacts on the reproductive system. J Women's Health 8(2):217-26.

Teitelbaum DT (1999). The toxicology of 1,2-dibromo-3-chloropropane (DBCP): a brief review. Int J Occup Environ Health 5(2):122-6.

Wiger R, Holme JA, Hongslo JK, Brunborg G, Haug K, Rodilla V, Dybing E, Soderlund EJ (1998). Single-strand breaks, cell cycle arrest and apoptosis in HL-60 and LLCPK1 cells exposed to 1,2-dibromo-3-chloropropane. Cell Biol Toxicol 14(4):267-82.

1,2-Dichlorobenzene

A PHG of 0.6 mg/L (600 ppb) was developed in 1997 for 1,2-dichlorobenzene (1,2-DCB, also known as ortho-dichlorobenzene) in drinking water. The PHG is based on several adverse effects, including hepatotoxicity and organ and body weight changes observed in experimental animals in a subchronic oral exposure study. 1,2-DCB is a solvent and a chemical intermediate. It also has been used as an insecticide/fumigant; the last

registered pesticide product containing 1,2-DCB as an active ingredient was removed from the California market in 1985.

1,2-DCB and other simple chlorinated benzenes have been used in the study of hepatocyte injury and repair mechanisms, which may provide some perspective on both the dose-response and structure-activity relationships for these chemicals (see also 1,4-DCB). Of particular interest is why 1,2-DCB causes hepatotoxicity but has not been found to cause liver tumors, while 1,4-DCB causes both hepatotoxicity and a clearly increased incidence of liver tumors.

References

Badger DA, Kuester RK, Sauer JM, Sipes IG (1997). Gadolinium chloride reduces cytochrome P450: relevance to chemical-induced hepatotoxicity. Toxicology 121(2):143-53.

Ban M, Hettich D, Goutet M, Binet S (1998). Serum-borne factor(s) of 1,1-dichloroethylene and 1,2-dichlorobenzene-treated mice inhibited in vitro antibody forming cell response and natural killer cell activity. Toxicol Lett 94(2):93-101.

Coenraads PJ, Tang NJ (2005). Chloracne. Contact Dermatitis 53(2):123.

Herr DW, Boyes WK (1997). A comparison of the acute neuroactive effects of dichloromethane, 1,3-dichloropropane, and 1,2-dichlorobenzene on rat flash evoked potentials (FEPs). Fundam Appl Toxicol 35(1):31-48.

Hissink AM, Oudshoorn MJ, Van Ommen B, Haenen GR, Van Bladeren PJ (1996). Differences in cytochrome P450-mediated biotransformation of 1,2-dichlorobenzene by rat and man: implications for human risk assessment. Chem Res Toxicol 9(8):1249-56.

Hissink AM, Van Ommen B, Kruse J, Van Bladeren PJ (1997). A physiologically based pharmacokinetic (PB-PK) model for 1,2-dichlorobenzene linked to two possible parameters of toxicity. Toxicol Appl Pharmacol 145(2):301-10.

Hissink AM, Van Ommen B, Van Bladeren PJ (1996). Dose-dependent kinetics and metabolism of 1,2-dichlorobenzene in rat: effect of pretreatment with phenobarbital. Xenobiotica 26(1):89-105.

Hoglen NC, Younis HS, Hartley DP, Gunawardhana L, Lantz RC, Sipes IG (1998). 1,2-Dichlorobenzene-induced lipid peroxidation in male Fischer 344 rats is Kupffer cell dependent. Toxicol Sci 46(2):376-85.

IARC (1999). Dichlorobenzenes. IARC Monogr Eval Carcinog Risks Hum 73:223-76.

Kulkarni SG, Duong H, Gomila R, Mehendale HM (1996). Strain differences in tissue repair response to 1,2-dichlorobenzene. Arch Toxicol 70(11):714-23.

Kulkarni SG, Harris AJ, Casciano DA, Mehendale HM. Differential protooncogene expression in Sprague Dawley and Fischer 344 rats during 1,2-dichlorobenzene-induced hepatocellular regeneration. Toxicology. 1999 Nov 29;139(1-2):119-27.

Lake BG, Cunninghame ME, Price RJ (1997). Comparison of the hepatic and renal effects of 1,4-dichlorobenzene in the rat and mouse. Fundam Appl Toxicol 39(1):67-75.

Nedelcheva V, Gut I, Soucek P, Frantik E (1998). Cytochrome P450 catalyzed oxidation of monochlorobenzene, 1,2- and 1,4-dichlorobenzene in rat, mouse, and human liver microsomes. Chem Biol Interact 115(1):53-70.

Paolini M, Pozzetti L, Silingardi P, Della Croce C, Bronzetti G, Cantelli-Forti G (1998). Isolation of a novel metabolizing system enriched in phase-II enzymes for short-term genotoxicity bioassays. Mutat Res 13(3):205-17.

Umemura T, Saito M, Takagi A, Kurokawa Y (1996). Isomer-specific acute toxicity and cell proliferation in livers of B6C3F1 mice exposed to dichlorobenzene. Toxicol Appl Pharmacol 137(2):268-74.

van Wijk D, Thompson RS, De Rooij C, Garny V, Lecloux A, Kanne R (2004). 1,2-dichlorobenzene marine risk assessment with special reference to the OSPARCOM region: North Sea. Environ Monit Assess 97(1-3):87-102.

Versonnen BJ, Arijs K, Verslycke T, Lema W, Janssen CR (2003). In vitro and in vivo estrogenicity and toxicity of o-, m-, and p-dichlorobenzene. Environ Toxicol Chem 22(2):329-35.

Violante FS, Milani S, Malenchini G, Barbieri A (2005). Chloracne due to odichlorobenzene in a laboratory worker. Contact Dermatitis 52(2):108.

Violante FS, Milani S, Malenchini G, Barbieri A (2005). Reply to the comments by Coenraads and Tang 'Chloracne due to o-dichlorobenzene in a laboratory worker', Contact Dermatitis 2005: 52:108. Contact Dermatitis 2005 Jul;53(1):65.

Younis HS, Hoglen NC, Kuester RK, Gunawardhana L, Sipes IG (2000). 1,2-Dichlorobenzene-mediated hepatocellular oxidative stress in Fischer-344 and Sprague-Dawley rats. Toxicol Appl Pharmacol 163(2):141-8.

Younis HS, Parrish AR, Glenn Sipes I (2003). The role of hepatocellular oxidative stress in Kupffer cell activation during 1,2-dichlorobenzene-induced hepatotoxicity. Toxicol Sci 76(1):201-11.

1,4-Dichlorobenzene

A PHG of 0.006 mg/L (6 ppb) was developed in 1997 for 1,4-dichlorobenzene (1,4-DCB), also known as para-dichlorobenzene, in drinking water. The PHG is based on hepatocarcinogenic effects in both male and female mice observed in a chronic study conducted by the National Toxicology Program (NTP). 1,4-DCB is used mainly as a fumigant for the control of moths, molds, and mildews (mothballs), and as a space deodorant for toilets and refuse containers. It is also used as an intermediate in the production of other chemicals, in the control of tree-boring insects, and in the control of mold in tobacco seeds.

The general population is exposed to 1,4-DCB through breathing vapors from household products such as mothballs and toilet deodorizer blocks. Traces of 1,4 DCB have been found in foods such as meat, eggs, and honey. Acute human exposure to 1,4-dichlorobenzene in air results in irritation to the eyes, skin, and throat. Subchronic exposure to experimental animals caused liver effects (degeneration and necrosis), bone

marrow hypoplasia, and renal damage and necrosis at all levels of ingestion. Chronic exposure to 1,4 DCB in mice showed renal damage, liver toxicity, increases in heart, liver, and lung weights, and parathyroid hyperplasia among males and neuropathy in females.

References

Aiso S, Arito H, Nishizawa T, Nagano K, Yamamoto S, Matsushima T (2005). Thirteenweek inhalation toxicity of p-dichlorobenzene in mice and rats. J Occup Health 47(3):249-60.

Aiso S, Takeuchi T, Arito H, Nagano K, Yamamoto S, Matsushima T (2005). Carcinogenicity and chronic toxicity in mice and rats exposed by inhalation to paradichlorobenzene for two years. J Vet Med Sci 67(10):1019-29.

ATSDR (1998). Toxicological Profile for 1,4-Dichlorobenzene (Update). Agency for Toxic Substances and Disease Registry, Public Health Service, U.S. Department of Health and Human Services, Atlanta, GA..

Bahrami F, Brittebo EB, Bergman A, Larsson C, Brandt I (1999). Localization and comparative toxicity of methylsulfonyl-2,5- and 2,6-dichlorobenzene in the olfactory mucosa of mice. Toxicol Sci 49(1):116-23.

Barter JA, Sherman JH (1999). An evaluation of the carcinogenic hazard of 1,4-dichlorobenzene based on internationally recognized criteria. Regul Toxicol Pharmacol 29(1):64-79.

Bomhard EM, Schmidt U, Loser E (1998). Time course of enzyme induction in liver and kidneys and absorption, distribution and elimination of 1,4-dichlorobenzene in rats. Toxicology 131(2-3):73-91.

Canonero R, Campart GB, Mattioli F, Robbiano L, Martelli A (1997). Testing of p-dichlorobenzene and hexachlorobenzene for their ability to induce DNA damage and micronucleus formation in primary cultures of rat and human hepatocytes. Mutagenesis 12(1):35-9.

Gustafson DL, Coulson AL, Feng L, Pott WA, Thomas RS, Chubb LS, Saghir SA, Benjamin SA, Yang RS (1998). Use of a medium-term liver focus bioassay to assess the hepatocarcinogenicity of 1,2,4,5-tetrachlorobenzene and 1,4-dichlorobenzene. Cancer Lett 129(1):39-44.

Gustafson DL, Long ME, Thomas RS, Benjamin SA, Yang RS (2000). Comparative hepatocarcinogenicity of hexachlorobenzene, pentachlorobenzene, 1,2,4,5-tetrachlorobenzene, and 1,4-dichlorobenzene: application of a medium-term liver focus bioassay and molecular and cellular indices. Toxicol Sci 53(2):245-52.

Hasmall SC, Roberts RA (1997). Hepatic ploidy, nuclearity, and distribution of DNA synthesis: a comparison of nongenotoxic hepatocarcinogens with noncarcinogenic liver mitogens. Toxicol Appl Pharmacol 144(2):287-93.

Hissink AM, Oudshoorn MJ, Van Ommen B, Van Bladeren PJ (1997). Species and strain differences in the hepatic cytochrome P450-mediated biotransformation of 1,4-dichlorobenzene. Toxicol Appl Pharmacol 145(1):1-9.

James NH, Soames AR, Roberts RA (1998). Suppression of hepatocyte apoptosis and induction of DNA synthesis by the rat and mouse hepatocarcinogen diethylhexylphthalate (DEHP) and the mouse hepatocarcinogen 1,4-dichlorobenzene (DCB). Arch Toxicol 72(12):784-90.

Makita Y (2005). Effects of perinatal combined exposure to 1,4-dichlorobenzene and 1,1-dichloro-2, 2-bis (p-chlorophenyl) ethylene on rat male offspring. Basic Clin Pharmacol Toxicol 96(5):361-5.

Mally A, Chipman JK (2002). Non-genotoxic carcinogens: early effects on gap junctions, cell proliferation and apoptosis in the rat. Toxicology 180(3):233-48.

Muller M (2002). 1,4-Dichlorobenzene-Induced liver tumors in the mouse: evaluation of the role of chlorohydroquinones. Rev Environ Health 17(4):279-90.

NTP (2002). 1,4-Dichlorobenzene. National Toxicology Program. Rep Carcinog 10:84-5.

Robbiano L, Carrozzino R, Puglia CP, Corbu C, Brambilla G (1999). Correlation between induction of DNA fragmentation and micronuclei formation in kidney cells from rats and humans and tissue-specific carcinogenic activity. Toxicol Appl Pharmacol 161(2):153-9.

Sasaki YF, Izumiyama F, Nishidate E, Matsusaka N, Tsuda S (1997). Detection of rodent liver carcinogen genotoxicity by the alkaline single-cell gel electrophoresis (Comet) assay in multiple mouse organs (liver, lung, spleen, kidney, and bone marrow). Mutat Res 391(3):201-14.

Sherman JH, Nair RS, Steinmetz KL, Mirsalis JC, Nestmann ER, Barter JA (1998). Evaluation of unscheduled DNA synthesis (UDS) and replicative DNA synthesis (RDS) following treatment of rats and mice with p-dichlorobenzene. Teratog Carcinog Mutagen 18(6):309-18.

Tegethoff K, Herbold BA, Bomhard EM (2000). Investigations on the mutagenicity of 1,4-dichlorobenzene and its main metabolite 2,5-dichlorophenol in vivo and in vitro. Mutat Res 470(2):161-7.

Umemura T, Kodama Y, Kurokawa Y, Williams GM (2000). Lack of oxidative DNA damage or initiation of carcinogenesis in the kidneys of male F344 rats given subchronic exposure to p-dichlorobenzene (pDCB) at a carcinogenic dose. Arch Toxicol 74(1):54-9.

Umemura T, Saito M, Takagi A, Kurokawa Y (1996). Isomer-specific acute toxicity and cell proliferation in livers of B6C3F1 mice exposed to dichlorobenzene. Toxicol Appl Pharmacol 137(2):268-74.

Umemura T, Takada K, Schulz C, Gebhardt R, Kurokawa Y, Williams GM (1998). Cell proliferation in the livers of male mice and rats exposed to the carcinogen P-dichlorobenzene: evidence for thresholds. Drug Chem Toxicol 21(1):57-66.

Veraldi A, Costantini AS, Bolejack V, Miligi L, Vineis P, van Loveren H (2006). Immunotoxic effects of chemicals: A matrix for occupational and environmental epidemiological studies. Am J Ind Med 49(12):1046-55.

Versonnen BJ, Arijs K, Verslycke T, Lema W, Janssen CR (2003). In vitro and in vivo estrogenicity and toxicity of o-, m-, and p-dichlorobenzene. Environ Toxicol Chem 22(2):329-35.

Williams GM, Iatropoulos MJ (2002). Alteration of liver cell function and proliferation: differentiation between adaptation and toxicity. Toxicol Pathol 30(1):41-53.

Yoshida T, Andoh K, Kosaka H, Kumagai S, Matsunaga I, Akasaka S, Nakamura S, Oda H, Fukuhara M (2002). Inhalation toxicokinetics of p-dichlorobenzene and daily absorption and internal accumulation in chronic low-level exposure to humans. Arch Toxicol 76(5-6):306-15.

1,1-Dichloroethylene

A PHG of 0.01 mg/L ($10 \mu g/L$, or 10 ppb) was developed in 1999 for 1,1-dichloroethylene (1,1-DCE, also known as vinylidene chloride) in drinking water. 1,1-DCE is used principally for the production of polyvinylidene chloride polymers. 1,1-DCE does not occur naturally, but can be found in landfills as a result of the breakdown of polyvinylidene chloride products. Owing to its high vapor pressure, air releases are the largest sources of the chemical in the environment. Concentrations of 1 to 550 $\mu g/mL$ 1,1-DCE have been reported in surface waters near industrial sites.

The 1999 PHG was based on the most sensitive toxic endpoint, midzonal hepatocellular fatty changes in female rats at 50 ppm (approximately 9 mg/kg-day) and greater in their drinking water. There appears to be inadequate evidence to consider 1,1-DCE to be a carcinogen. Inhalation exposure to DCE is both fetotoxic and produces developmental effects in laboratory animal studies at maternally toxicity doses. Other recent studies on lung and immune system effects of 1,1-DCE deserve additional consideration.

References

Ban M, Hettich D, Bonnet P (2003). Effect of inhaled industrial chemicals on systemic and local immune response. Toxicology 184(1):41-50.

Ban M, Hettich D, Goutet M, Binet S (1998). Serum-borne factor(s) of 1,1-dichloroethylene and 1,2-dichlorobenzene-treated mice inhibited in vitro antibody forming cell response and natural killer cell activity. Toxicol Lett 94(2):93-101.

Ban M, Langonne I, Huguet N, Pepin E, Morel G (2006). Inhaled chemicals may enhance allergic airway inflammation in ovalbumin-sensitized mice. Toxicology 226(2-3):161-71.

Battershill JM, Fielder RJ (1998). Mouse-specific carcinogens: an assessment of hazard and significance for validation of short-term carcinogenicity bioassays in transgenic mice. Hum Exp Toxicol 17(4):193-205.

Dowsley TF, Reid K, Petsikas D, Ulreich JB, Fisher RL, Forkert PG (1999). Cytochrome P-450-dependent bioactivation of 1,1-dichloroethylene to a reactive epoxide in human lung and liver microsomes. J Pharmacol Exp Ther 289(2):641-8.

Forkert PG, Boyd SM, Ulreich JB (2001). Pulmonary bioactivation of 1,1-dichloroethylene is associated with CYP2E1 levels in A/J, CD-1, and C57BL/6 mice. J Pharmacol Exp Ther 297(3):1193-200.

Forkert PG, Boyd SM (2001). Differential metabolism of 1,1-dichloroethylene in livers of A/J, CD-1, and C57BL/6 mice. Drug Metab Dispos 29(11):1396-402.

Forkert PG (1999). 1,1-Dichloroethylene-induced Clara cell damage is associated with in situ formation of the reactive epoxide. Immunohistochemical detection of its glutathione conjugate. Am J Respir Cell Mol Biol 20(6):1310-8.

Forkert PG (1998). Immunohistochemical detection of CYP2E1 and 2-S-glutathionyl acetate in murine lung tumors: diminished formation of reactive intermediates of 1,1-dichloroethylene. Exp Lung Res 24(4):455-61.

Forkert PG (2001). Mechanisms of 1,1-dichloroethylene-induced cytotoxicity in lung and liver. Drug Metab Rev 33(1):49-80.

Gram TE (1997). Chemically reactive intermediates and pulmonary xenobiotic toxicity. Pharmacol Rev 49(4):297-341.

IARC (1999). Vinylidene chloride. IARC Monogr Eval Carcinog Risks Hum 71 Pt 3:1163-80.

Jones JA, Kaphalia L, Treinen-Moslen M, Liebler DC (2003). Proteomic characterization of metabolites, protein adducts, and biliary proteins in rats exposed to 1,1-dichloroethylene or diclofenac. Chem Res Toxicol 16(10):1306-17.

Martin EJ, Forkert PG (2005). 1,1-Dichloroethylene-induced mitochondrial damage precedes apoptotic cell death of bronchiolar epithelial cells in murine lung. J Pharmacol Exp Ther 313(1):95-103.

Martin EJ, Racz WJ, Forkert PG (2003). Mitochondrial dysfunction is an early manifestation of 1,1-dichloroethylene-induced hepatotoxicity in mice. J Pharmacol Exp Ther 304(1):121-9.

Roberts SM, Jordan KE, Warren DA, Britt JK, James RC (2002). Evaluation of the carcinogenicity of 1,1-dichloroethylene (vinylidene chloride). Regul Toxicol Pharmacol 35(1):44-55.

Simmonds AC, Reilly CA, Baldwin RM, Ghanayem BI, Lanza DL, Yost GS, Collins KS, Forkert PG (2004). Bioactivation of 1,1-dichloroethylene to its epoxide by CYP2E1 and CYP2F enzymes. Drug Metab Dispos 32(9):1032-9.

Slikker W Jr, Andersen ME, Bogdanffy MS, Bus JS, *et al.* (2004). Dose-dependent transitions in mechanisms of toxicity: case studies. Toxicol Appl Pharmacol 201(3):226-94.

Slikker W Jr, Andersen ME, Bogdanffy MS, Bus JS, et al. (2004). Dose-dependent transitions in mechanisms of toxicity. Toxicol Appl Pharmacol 201(3):203-25.

U.S. EPA (2002). Health assessment documents for vinylidene chloride: Updated Final Report. U.S. Environmental Protection Agency, Washington, DC.

Warbrick EV, Dearman RJ, Ashby J, Schmezer P, Kimber I (2001). Preliminary assessment of the skin sensitizing activity of selected rodent carcinogens using the local lymph node assay. Toxicology 163(1):63-9.

Williams PR, Patterson J, Briggs DW (2006). VCCEP pilot: progress on evaluating children's risks and data needs. Risk Anal 26(3):781-801.

Woodard SH, Moslen MT (1998). Decreased biliary secretion of proteins and phospholipids by rats with 1,1-dichloroethylene-induced bile canalicular injury. Toxicol Appl Pharmacol 152(2):295-301.

1,2-Dichloropropane

A PHG of 0.5 μ g/L (or ppb) was developed in 1999 for 1,2-dichloropropane (1,2-DCP) in drinking water, based on carcinogenic effects observed in experimental animals. 1,2-DCP, also known as propylene dichloride, is primarily used as a chemical intermediate in the synthesis of other chlorinated hydrocarbons. In the past, it was used as a fumigant and industrial solvent. It may be found as a contaminant in the fumigant Telone (1,3-dichloropropene).

Kidney and liver damage has been observed after acute and subchronic exposure to 1,2-DCP, both in humans and in experimental animals. Tests for mutagenicity and genotoxicity have been mixed, some negative, some positive. 1,2-DCP has been frequently detected at low levels in air as well as ground and surface water, although levels should have decreased due to improved industrial vapor controls and discontinuation of its use as a fumigant. The recent research on reproductive system effects of 1,2-DCP in female rats particularly deserves review.

References

Chroust K, Pavlova M, Prokop Z, Mendel J, Bozkova K, Kubat Z, Zajickova V, Damborsky J (2007). Quantitative structure-activity relationships for toxicity and genotoxicity of halogenated aliphatic compounds: wing spot test of Drosophila melanogaster. Chemosphere 67(1):152-9.

Fiaccadori E, Maggiore U, Rotelli C, Giacosa R, Ardissino D, De Palma G, Bergamaschi E, Mutti A (2003). Acute renal and hepatic failure due to accidental percutaneous absorption of 1,2-dichlorpropane contained in a commercial paint fixative. Nephrol Dial Transplant 18(1):219-20.

Heindel JJ, Chapin RE, Gulati DK, George JD, Price CJ, Marr MC, Myers CB, Barnes LH, Fail PA, Grizzle TB, *et al.* (1994). Assessment of the reproductive and developmental toxicity of pesticide/fertilizer mixtures based on confirmed pesticide contamination in California and Iowa groundwater. Fundam Appl Toxicol 22(4):605-21.

IARC (1999). 1,2-Dichloropropane. IARC Monogr Eval Carcinog Risks Hum 71 Pt 3:1393-400.

Lazo-Torres AM, Lopez-Caler C, Galvez-Contreras C, Yelamos-Rodriguez F (2005). [Dichloropropane poisoning: report of 2 cases] Med Clin (Barc) 124(8):318.

Lefever MR, Wackett LP (1994). Oxidation of low molecular weight chloroalkanes by cytochrome P450CAM. Biochem Biophys Res Commun 201(1):373-8.

Lucantoni C, Grottoli S, Gaetti R (1992). 1,2-Dichloropropane is a renal and liver toxicant. Toxicol Appl Pharmacol 117(1):133.

Odinecs A, Maso S, Nicoletto G, Secondin L, Trevisan A (1995). Mechanism of sexrelated differences in nephrotoxicity of 1.2-dichloropropane in rats. Ren Fail 17(5):517-24.

Sekiguchi S, Suda M, Zhai YL, Honma T (2002). Effects of 1-bromopropane, 2-bromopropane, and 1,2-dichloropropane on the estrous cycle and ovulation in F344 rats. Toxicol Lett 126(1):41-9.

Trevisan A, Meneghetti P, Maso S, Secondin L, Nicoletto G (1992). Sex- and age-related nephrotoxicity due to 1,2-dichloropropane in vitro. Arch Toxicol 66(9):641-5.

Trevisan A, Meneghetti P, Maso S, Troso O (1993). In-vitro mechanisms of 1,2-dichloropropane nephrotoxicity using the renal cortical slice model. Hum Exp Toxicol 12(2):117-21.

Vitali M, Ensabella F, Stella D, Guidotti M (2006). Exposure to organic solvents among handicraft car painters: A pilot study in Italy. Ind Health 44(2):310-7.

Yang R (1993). NTP technical report on the toxicity studies of Pesticide/Fertilizer Mixtures Administered in Drinking Water to F344/N Rats and B6C3F1 Mice. Toxic Rep Ser. 36:1-G3.

Dinoseb

A PHG of 14 ppb was developed for dinoseb (2-sec-butyl-4,6-dinitrophenol) in drinking water in 1997. Dinoseb is a dinitrophenolic compound once used extensively as an herbicide and pesticide in California. It was suspended for all pesticide uses by U.S. EPA in 1986 based on concern about potential reproductive and teratogenic effects in agricultural workers. It has not been detected in California water supplies in several years.

Dinoseb, like other dinitrophenols, is an inhibitor of mitochondrial respiration (the source of most cellular energy, used to support metabolism). More recent studies of this effect have provided new perspectives on its spermatocidal and teratogenic effects.

References

Branch S, Rogers JM, Brownie CF, Chernoff N (1996). Supernumerary lumbar rib: manifestation of basic alteration in embryonic development of ribs. J Appl Toxicol 16(2):115-9.

Kato M, Fukunishi K, Ikegawa S, Higuchi H, Sato M, Horimoto M, Ito S (2001). Overview of studies on rat sperm motion analysis using a Hamilton-Thorne Sperm Analyzer--collaborative working study. J Toxicol Sci 26(5):285-97.

Palmeira CM, Moreno AJ, Madeira VM (1994). Interactions of herbicides 2,4-D and dinoseb with liver mitochondrial bioenergetics. Toxicol Appl Pharmacol 127(1):50-7.

Palmeira CM, Moreno AJ, Madeira VM (1995). Effects of paraquat, dinoseb and 2,4-D on intracellular calcium and on vasopressin-induced calcium mobilization in isolated hepatocytes. Arch Toxicol 69(7):460-6.

Palmeira CM, Moreno AJ, Madeira VM (1995). Thiols metabolism is altered by the herbicides paraquat, dinoseb and 2,4-D: a study in isolated hepatocytes. Toxicol Lett 81(2-3):115-23.

Rogers JM, Setzer RW, Branch S, Chernoff N (2004). Chemically induced supernumerary lumbar ribs in CD-1 mice: size distribution and dose response. Birth Defects Res B Dev Reprod Toxicol 71(1):17-25.

Takahashi KL, Aoyama H, Kawashima K, Teramoto S (2003). Effects of dinoseb, 4,6-dinitro-o-cresol, and 2,4-dinitrophenol on rat Sertoli-germ cell co-cultures. Reprod Toxicol 17(2):247-52.

Takahashi KL, Hojo H, Aoyama H, Teramoto S (2004). Comparative studies on the spermatotoxic effects of dinoseb and its structurally related chemicals. Reprod Toxicol 18(4):581-8.

Endothall

A PHG of 580 ppb for endothall (7-oxabicyclo(2.2.1) heptane-2,3 dicarboxylic acid) in drinking water was developed in 1997 based on gastric toxicity observed in dogs. Endothall is an organic acid effective as a contact weed killer. Major uses include the defoliation of cotton, control of algae and aquatic weeds and as a desiccant on other crops. Its selective action and rapid breakdown to nontoxic products makes it desirable for control of aquatic weeds, leaving other desirable species such as fish and insects relatively unaffected.

As a strong organic acid, endothall is poorly absorbed both orally and dermally, and its effects tend to be limited to direct irritancy. However, endothall is considered to be a selective inhibitor of type 2A protein phosphatase (PP2A(c)), and has been used for this purpose in metabolic studies. The recent reevaluation of endothall for reregistration by U.S. EPA (2005) included extra consideration of exposures for infants and children, and concluded that drinking water exposure for infants less than one year old is at the level of concern (using the U.S. EPA MCL of 100 ppb as a surrogate for actual drinking water exposure data). OEHHA will evaluate this conclusion in the context of our mandate for protecting sensitive populations.

References

Boknik P, Vahlensieck U, Huke S, Knapp J, Linck B, Luss H, Muller FU, Neumann J, Schmitz W (2000). On the cardiac contractile, electrophysiological and biochemical effects of endothall, a protein phosphatase inhibitor. Pharmacology 61(1):43-50.

Erdodi F, Toth B, Hirano K, Hirano M, Hartshorne DJ, Gergely P (1995). Endothall thioanhydride inhibits protein phosphatases-1 and -2A in vivo. Am J Physiol 269(5 Pt 1):C1176-84.

Thiery JP, Blazsek I, Legras S, Marion S, Reynes M, Anjo A, Adam R, Misset JL (1999). Hepatocellular carcinoma cell lines from diethylnitrosamine phenobarbital-treated rats. Characterization and sensitivity to endothall, a protein serine/threonine phosphatase-2A inhibitor. Hepatology 29(5):1406-17.

U.S. EPA (2005). Reregistration Eligibility Decision for Endothall. Prevention, Pesticides and Toxic Substances, Environmental Protection Agency, Washington, DC. EPA 738-R-05-008. Accessed at: http://www.epa.gov/REDs/endothall_red.pdf.

Wnorowski G (1997). Acute Dermal Toxicity Limit Test (in Rabbits): ThinRite: Lab Project Number: 5072: P322. Unpublished study prepared by Product Safety Labs. 16 p. U.S. EPA MRID 44319606.03-Jul-1997.

Wnorowski G (1997). Acute Inhalation Toxicity Limit Test (in Rats): ThinRite: Lab Project Number: 5076: P330. Unpublished study prepared by Product Safety Labs. 24 p. U.S. EPA MRID 44319607. 03-Jul-1997.

Wnorowski G (1997). Acute Oral Toxicity Defined LD50 (in Rats): ThinRite: Lab Project Number: 5071: P320. Unpublished study prepared by Product Safety Labs. 25 p. U.S. EPA MRID 44319605. 03-Jul-1997.

Wnorowski G (1997). Primary Eye Irritation (in Rabbits): ThinRite: Lab Project Number: 5073: P324. Unpublished study prepared by Product Safety Labs. 22 p. U.S. EPA MRID 44319608. 03-Jul-1997.

Wnorowski G (1997). Dermal Sensitization Test--Buehler Method (in Guinea Pigs): ThinRite: Lab Project Number: 5075: P328. Unpublished study prepared by Product Safety Labs. 24 p. U.S. EPA MRID 44319610. 03-Jul-1997.

Wnorowski G (1997). Primary Skin Irritation (in Rabbits): ThinRite: Lab Project Number: 5074: P326. Unpublished study prepared by Product Safety Labs. 17 p. U.S. EPA MRID 44319609. 03-Jul-1997.

Endrin

A PHG of 0.0018 mg/L (1.8 ppb) was developed for endrin in drinking water in 1999. The PHG is based on the observation of seizures and minor pathological changes to the liver in dogs fed 2 and 4 ppm endrin for two years. Endrin is an environmentally persistent chlorinated hydrocarbon pesticide similar to aldrin and dieldrin, which are classified as carcinogens. It has been postulated that the failure to demonstrate carcinogenicity of the structurally related compound endrin is that the high toxicity of endrin limits the daily dose to levels at which the tumor incidence would be insignificant. The biological half-life of endrin in mammals is also quite short, compared to chlorinated hydrocarbons like dieldrin and DDT. Mechanistic data that might help resolve the question of its potential carcinogenicity has not been produced since the publication of the earlier PHG.

Most uses of endrin were cancelled by the U.S. EPA in 1979, and the remaining limited usage was cancelled in 1991, so there has been little incentive for further toxicity studies. Endrin residues persist in the environment in aqueous sediments, soil, and marine fish, but not in red meat and poultry.

References

Bagchi D, Balmoori J, Bagchi M, Ye X, Williams CB, Stohs SJ (2000). Role of p53 tumor suppressor gene in the toxicity of TCDD, endrin, naphthalene, and chromium (VI) in liver and brain tissues of mice. Free Radic Biol Med 28(6):895-903.

Bagchi D, Balmoori J, Bagchi M, Ye X, Williams CB, Stohs SJ (2002). Comparative effects of TCDD, endrin, naphthalene and chromium (VI) on oxidative stress and tissue damage in the liver and brain tissues of mice. Toxicology 175(1-3):73-82.

Carr RL, Couch TA, Liu J, Coats JR, Chambers JE (1999). The interaction of chlorinated alicyclic insecticides with brain GABA(A) receptors in channel catfish (Ictalurus punctatus). J Toxicol Environ Health A 56(8):543-53.

Hassoun EA, Stohs SJ (1996). Comparative teratological studies on TCDD, endrin and lindane in C57BL/6J and DBA/2J mice. Comp Biochem Physiol C Pharmacol Toxicol Endocrinol 113(3):393-8.

Hassoun EA, Stohs SJ (1996). TCDD, endrin and lindane induced oxidative stress in fetal and placental tissues of C57BL/6J and DBA/2J mice. Comp Biochem Physiol C Pharmacol Toxicol Endocrinol 115(1):11-8.

Huang X, Hites RA, Foran JA, Hamilton C, Knuth BA, Schwager SJ, Carpenter DO (2006). Consumption advisories for salmon based on risk of cancer and noncancer health effects. Environ Res 101(2):263-74.

Khan MA, Jovanovich LV, Martin LT, Qadri SY, Podowski AA (1998). Effects of photoisomers of cyclodiene insecticides on liver and microsomal cytochrome P450 in rats. Arch Toxicol 72(2):74-83.

Lemaire G, Balaguer P, Michel S, Rahmani R (2005). Activation of retinoic acid receptor-dependent transcription by organochlorine pesticides. Toxicol Appl Pharmacol 202(1):38-49.

Mumtaz MM, Tully DB, El-Masri HA, De Rosa CT (2002). Gene induction studies and toxicity of chemical mixtures. Environ Health Perspect 110 Suppl 6:947-56.

Weston DP, You J, Lydy MJ (2004). Distribution and toxicity of sediment-associated pesticides in agriculture-dominated water bodies of California's Central Valley. Environ Sci Technol 38(10):2752-9.

Hexachlorocyclopentadiene

A PHG of 0.05 mg/L (50 ppb) was developed in 1999 for hexachlorocyclopentadiene (HCCPD) in drinking water, based on stomach lesions after oral administration of HCCPD to rats and mice. HCCPD is a halogenated hydrocarbon used as an intermediate in the production of dyes, resins, pharmaceuticals, flame retardants, and insecticides. HCCPD is also used in the production of a variety of industrial chemicals, including ketones, fluorocarbons, acids, esters, and plastics. The major toxic effect of HCCPD is irritation at the point of contact (lungs, skin, or stomach). Kidney and liver damage also occurs at moderate doses after oral (gavage) administration to rodents.

Our preliminary literature scan revealed no new toxicity studies on this chemical. The updated U.S. EPA review takes an approach similar to the 1987 U.S. EPA review (and the 1999 PHG), except for a benchmark calculation of the reference dose.

References

ATSDR (1999). Toxicological profile for hexachlorocyclopentadiene (HCCPD). Agency for Toxic Substances and Disease Registry, Public Health Service, U.S. Dept. of Health and Human Services, Atlanta, GA. July 1999. Accessed at: www.atsdr.cdc.gov/toxprofiles/tp112.pdf.

U.S. EPA (2001). Toxicological review of hexachlorocyclopentadiene (CAS No. 77-47-4) U.S. Environmental Protection Agency Washington, DC. June 2001. EPA/600/R-01/013. Accessed at: www.epa.gov/iris/toxreviews/0059-tr.pdf.

U.S. EPA (2001b). Hexachlorocyclopentadiene (HCCPD) (CASRN 77-47-4), last revised 07/05/2001. Integrated Risk Information System, U.S. Environmental Protection Agency, Washington, DC. Accessed at: http://www.epa.gov/iris/subst/0059.htm.

Methoxychlor

In 1999, OEHHA published a PHG of 0.03 mg/L (30 ppb) for methoxychlor (MXC) in drinking water. MXC is a chlorinated hydrocarbon insecticide that is approved for use on livestock as well as on numerous agricultural crops. MXC was suspended from use in California in 1995 and in the rest of the country in 2000; U.S. EPA revoked all tolerances in crops in 2002. MXC has been found in surface waters near points of application for pest control, and in groundwater near waste disposal sites; it has not been detected in California municipal drinking water. Very high doses of MXC can cause tremors. convulsions, and other signs of neurological stimulation. The most significant effects of repeated exposure to MXC are on reproductive tissues; both the parent compound and its metabolites exhibit estrogenic activity. Chronic and sub-chronic exposure can produce adverse effects on the male and female reproductive system, developmental effects, and long-term effects on neurobehavioral development. Carcinogenicity studies on MXC are inadequate; both the U.S. EPA and IARC have judged MXC to be not classifiable as to human carcinogenicity. The PHG for MXC is based on reproductive effects in female rats exposed during the perinatal period. The data are supported by similar findings in other reproductive and developmental studies in animals. No human studies on human sensitivity to the potential endocrine-disruptive effects of this pesticide are available.

A large volume of new literature, predominantly relating to the estrogenic effects of MXC and its metabolites in animal studies, has been published since the PHG was developed in 1999. Several new mechanism studies are also available. Reported reproductive tract abnormalities at doses lower than those used to derive the original PHG also deserve intensive review. Representative new studies available are listed below.

References

Alworth LC, Howdeshell KL, Ruhlen RL, Day JK, et al. (2002). Uterine responsiveness to estradiol and DNA methylation are altered by fetal exposure

to diethylstilbestrol and methoxychlor in CD-1 mice: effects of low versus high doses. Toxicol Appl Pharmacol 183:10-22.

Eroschenko VP, Amstislavsky SY, Schwabel H, Ingermann RL (2002). Altered behaviors in male mice, male quail, and salamander larvae following early exposures to the estrogenic pesticide methoxychlor. Neurotoxicol Teratol 24:29-36.

Gioiosa L, Fissore E, Ghirardelli G, *et al.* (2007). Developmental exposure to low-dose estrogenic endocrine disruptors alters sex differences in exploration and emotional responses in mice. Horm Behav May 22 [Epub ahead of print].

Golub MS, Germann SL, Hogrefe CE (2004). Endocrine disruption and cognitive function in adolescent female rhesus monkeys. Neurotox Teratol 26(6):799-809.

Golub MS, Hogrefe CE, Germann SL, Lasley BL, Natarajan K, Tarantal AF (2003). Effects of exogenous estrogenic agents on pubertal growth and reproductive system maturation in female rhesus monkeys. Toxicol Sci 74(1):103-13.

Golub MS, Hogrefe CE, Germann SL, *et al.* (2004). Endocrine disruption in adolescence: immunologic, hematologic and bone effects in monkeys. Toxicol Sci 82(2):598-607.

Gupta RK, Schuh RA, Fiskum G, Flaws JA (2006). Methoxychlor causes mitochondrial dysfunction and oxidative damage in the mouse ovary. Toxicol Appl Pharmacol 216(3):436-45.

Judy BM, Nagel SC, Thayer KA, Vom Saal FS, Welshons WV (1999). Low-dose bioactivity of xenoestrogens in animals: fetal exposure to low doses of methoxychlor and other xenoestrogens increases adult prostate size in mice. Toxicol Indus Health 15(1-2):12-25.

Latchoumycandane C, Chitra KC, Mathur PP (2002). The effect of methoxychlor on the epididymal antioxidant system of adult rats. Reprod Toxicol 16:161-72.

Latchoumycandane C, Mathur PP (2002). Induction of oxidative stress in the rat testis after short-term exposure to the organochlorine pesticide methoxychlor. Arch Toxicol 76:692-8.

Lopez–Espinosa MJ, Granada A, Carreno J, *et al.* (2007). Organochlorine pesticides in placentas from southern Spain and some related factors. Placenta 28(7):631-8.

Madhabananda LY, Bartolucci E, McIntyre BS, *et al.* (2002). Modulation of mammary gland development in prepubertal male rats exposed to genistein and methoxychlor. Toxicol Sci 66:216-25.

Matsutomi N, Shibutani M, Takagi H, *et al.* (2003). Impact of dietary exposure to methoxychlor, genistein, or diisononyl phthalate during the perinatal period on the development of the rat endocrine/reproductive systems in later life. Toxicol 192:149-170.

Oropeza-Hernandez L, Lopez-Romero R, Albores A (2003). Hepatic CYP1A, 2B, 2C, 2E and 3A regulation by methoxychlor in male and female rats. Toxicol Lett 144:93-103.

Palanza P, Morellini F, *et al.* (2002). Ethological methods to study the effects of maternal exposure to estrogenic endocrine disruptors: A study with methoxychlor. Neurotoxicol Teratol 24:55-69.

Savabieasfahani M, Kannan K, Astapova O, Evans NP, Padmanabhan V (2006). Developmental programming: differential effects of prenatal exposure to bisphenol-A or methoxychlor on reproductive function. Endocrinol 147(12):5956-66.

Shin JH, Moon HJ, Kang IH, Kim TS, *et al.* (2007). Calbindin-D9k mRNA expression in the rat uterus following exposure to methoxychlor: a comparison of oral and subcutaneous exposure. J Reprod Toxicol 53(2):179-88.

Smialowicz RJ (2002). The rat as a model in developmental immunotoxicology. Hum Exp Toxicol 21:513-19.

Staub C, Hardy V, Chapin R, Harris MW, Johnson L (2002). The hidden effect of estrogenic/antiandrogenic methoxychlor on spermatogenesis. Toxicol Appl Pharmacol 180:129-35.

Stoker T, Robinette CL, Cooper RL (1999). Perinatal exposure to estrogenic compounds and the subsequent effects on the prostate of the adult rat. Reprod Toxicol 13:463-72.

Swartz W, Eroschenko V (1998). Neonatal exposure to technical methoxychlor alters pregnancy outcome in female mice. Reprod Toxicol 12(6):565-73.

Takagi H, Mitsumori K, *et al.* (2002). Improvement of a two-stage carcinogenesis model to detect modifying effects of endocrine disrupting chemicals on thyroid carcinogenesis in rats. Cancer Lett 178(1):1-9.

Thompson S, Vorster SJ (2000). Attempted suicide by ingestion of methoxychlor. J Anal Toxicol 25(5):377-80.

Uzumcu M, Kuhn PE, Marano JE, *et al.* (2006). Early postnatal methoxychlor exposure inhibits folliculogenesis and stimulates anti-Mullerian hormone production in the rat ovary. Endocrinol 191(3):549-58.

Wang XJ, Bartolucci-Page E, Fenton SE, You L (2006). Altered mammary gland development in male rats exposed to genistein and methoxychlor. Toxicol Sci 91(1):93-103.

Takeuchi Y, Kosaka T, Hayashi K, Takeda M, Yoshida T, Fujisawa H, Teramoto S, Maita K, Harada T (2002). Thymic atrophy induced by methoxychlor in rat pups. Toxicol Lett 135:199-207.

U.S. EPA (2004). Methoxychlor Reregistration Eligibility Decision (RED). June 30, 2004. EPA Publication No. EPA 738-R-04-010. U.S. Environmental Protection Agency, Washington, DC. Accessed at: www.epa.gov/oppsrrd1/REDs/methoxychlor_red.htm.

Oxamyl

A PHG of 50 ppb was developed for oxamyl (S-methyl N'N'-dimethyl-N-[(methylcarbamoyl) oxy]-1-thiooxamimidate) in drinking water in 1997. Oxamyl is a carbamate insecticide which acts by inhibition of the enzyme acetylcholinesterase. Its

acute toxicity is very high. The risk assessment is based on decreased body weight gain, presumably caused by the ill effects of cholinesterase inhibition. Oxamyl is readily degraded in soil and groundwater; it has rarely been detected in California water sources, although it has been reported at low levels in drinking water from several other states.

The inhibition of cholinesterase caused by carbamates is very short-acting, so acute effects are most relevant. New studies by Malley *et al.* (1997, 1998) involving gavage administration of oxamyl to rats appear to provide a much lower no-observed-adverse-effect level than previous studies, and are likely to lead to a decreased PHG. The calculation should be based on exposure of an infant or toddler, because these age groups consume much greater amounts of water per body weight equivalent than do adults, and thus represent a susceptible population.

References

EWG (2007). National Contaminant Report: Oxamyl (Vydate). National Tap Water Quality Database. Environmental Working Group, Washington, D.C. Accessed at: http://www.ewg.org/tapwater/contaminants/contaminant.php?contamcode=2036.

Malley LA (1997). Acute oral neurotoxicity study of oxamyl technical in rats: Lab Project Number: HLR 1118-96: 10730-001. April, 1997. du Pont Haskell Laboratory for Toxicology and Industrial Medicine.

Malley LA (1998). Oxamyl technical: subchronic oral neurotoxicity study in rats: Lab Project Number: 10730: HL-1998-00708. du Pont Haskell Laboratory for Toxicology and Industrial Medicine.

U.S. EPA (2000a). Interim Reregistration Eligibility Decision (IRED): Oxamyl. Office of Prevention, Pesticides and Toxic Substances, U.S. Environmental Protection Agency, Washington, DC. EPA-738-R-00-015.

U.S. EPA (2000b). National Primary Drinking Water Regulations: Announcement of the Results of EPA's Review of Existing Drinking Water Standards and Request for Public Comment; Proposed Rule. U.S. Environmental Protection Agency. Fed Reg 67(74):19076-19078. April 17.

U.S. EPA (2000c). Memorandum: Oxamyl: amended toxicology chapter for RED. Office of Prevention, Pesticides and Toxic Substances, U.S. Environmental Protection Agency, Washington, DC. July 25, 2000.

U.S. EPA (2000d). Data Evaluation Report, Oxamyl, Chronic (1-Year) Toxicity Study - Dog (83-1b). Office of Pesticide Programs, U.S. Environmental Protection Agency, Washington, DC.

U.S. EPA (2004). Drinking water health advisory for oxamyl. Health and Ecological Criteria Division, Office of Water, U.S. Environmental Protection Agency, Washington D.C. September 2004. EPA-822-B-04-002.

Van Pelt CS (1999). DuPont's position on the NOEL in male dogs following chronic dietary exposure to oxamyl. DuPont Reports HLR 381-90 and HLO 555-90. Laboratory Project ID DuPont-2019. January 18, 1999. U.S. EPA MRID 44737503.

Pentachlorophenol

The PHG for pentachlorophenol (PCP) of 0.4 parts per billion was developed in 1997, and is based on carcinogenic effects in mice. PCP has primarily been used as a pesticide, for wood treatment and as a disinfectant. Wood used for decks, railings, and playground structures was frequently treated with PCP. Widespread human exposure occurred from dermal and inhalation exposures, as well as hand-to-mouth contact by children. PCP also leached into surface and groundwater. Because of its hazards, PCP is now restricted to use in a heat and pressure wood treatment process, for special outdoor applications such as utility poles.

In animal studies, short-term to chronic treatment with PCP has been reported to cause adverse effects on kidney and liver, increased fetal reabsorptions, anemia and leukopenia, and thyroid hormone disruption at relatively low levels (NOAELs <10 mg/kg-day). Significant increases in tumors in mice occur at about 20 mg/kg-day or more. A relatively large number of toxicity studies have been published since the release of the original PHG document in 1997. Major toxicity reviews have been published by ATSDR and U.S. EPA. OEHHA has also reviewed PCP for development of a child-specific reference dose for school siting and a No Significant Risk Level for Proposition 65.

References

ATSDR (2001). Toxicological Profile for Pentachlorophenol. Agency for Toxic Substances and Disease Registry, U.S. Public Health Service. Accessed 5/07 at http://www.atsdr.cdc.gov/toxprofiles/tp51.html.

ATSDR (2004). Airborne Chemicals from Wood Treatment Chemical. Agency for Toxic Substances and Disease Registry, U.S. Public Health Service. Accessed 5/07 at http://www.atsdr.cdc.gov/HAC/pha/MeredithCWilliam092904-GA/MeredithCWilliams092904HC-GA.pdf.

Beard A, Rawlings N (1998). Reproductive effects in mink (mustela vison) exposed to the pesticides lindane, carbofuran and pentachlorophenol in a multigeneration study. J Reprod Fertil 113:95-104.

Beard A, Rawlings N (1999). Thyroid function and effects on reproduction in ewes exposed to the organochlorine pesticides lindane or pentachlorophenol (PCP) from conception. J Toxicol Environ Health, Part A 58:509-530.

Bernard B, Hoberman A (2001). A study of the developmental toxicity potential of pentachlorophenol in the rat. Int J Toxicol 20:353-362.

Bernard B, Ranpuria A, Hoberman A (2001). Developmental toxicity study of pentachlorophenol in the rabbit. Int J Toxicol 20:345-352.

Bernard B, Hoberman A, Brown W, Ranpuria A, Christian M (2002). Oral (gavage) two-generation (one litter per generation) reproduction study of pentachlorophenol (penta) in rats. Int J Toxicol 21:301-318.

Blakley B, Yole M, Brousseau P, Boermans H, Fournier M (1998). Effect of pentachlorophenol on immune function. Toxicology 125:141-148.

CDPR (1998). Pentachlorophenol Risk Characterization Document. California Department of Pesticide Regulation, California Environmental Protection Agency, Sacramento, CA. Accessed at www.cdpr.ca.gov/docs/risk/rcd.htm.

Chhabra R, Maronpot R, Bucher J, Haseman J, Toft J, Hejtmancik M (1999). Toxicology and carcinogenesis studies of pentachlorophenol in rats. Toxicol Sci 48:14-20.

Daniel V, Huber W, Bauer K, Suesal C, Mytilineos J, Melk A, Conradt C, Opelz G (2001). Association of elevated blood levels of pentachlorophenol (PCP) with cellular and humoral immunodeficiencies. Arch Environ Health 56:77-83.

Danzo B, Shappell H, Banerjee A, Hachey D (2002). Effects of nonylphenol, 1,1-dichloro-2,2-bis(p-chlorophenyl)ethylene (p,p'-DDE), and pentachlorophenol on the adult female guinea pig reproductive tract. Reprod Toxicol 16:29-43.

Demers P, Davies H, Friesen M, Hertzman C, Ostry A, Hershler R, Teschke K (2006). Cancer and occupational exposure to pentachlorophenol and tetrachlorophenol (Canada). Cancer Causes Control 17:749-758.

Gerhard I, Daniel V, Link S, Monga B, Runnebaum B (1998). Chlorinated hydrocarbons in women with repeated miscarriages. Environ Health Perspect 106:675-681.

Gerhard I, Frick A, Monga B, Runnebaum B (1999). Pentachlorophenol exposure in women with gynecological and endocrine dysfunction. Environ Res Sec A 80:383-388.

Jung J, Ishida K, Nishihara T (2004). Anti-estrogenic activity of fifty chemicals evaluated by *in vitro* assays. Life Sci 74:3065-3074.

Lemaire G, Mnif W, Mauvais P, Balaguer P, Rahmani R (2006). Activation of α - and β -estrogen receptors by persistent pesticides in reporter cell lines. Life Sci 79:1160-1169.

Morville S, Scheyer A, Mirabel P, Millet M (2006). Spatial and geographical variations of urban, suburban and rural atmospheric concentrations of phenols and nitrophenols. Environ Sci Pollut Res 13:83-89.

NTP (1999). Toxicology and carcinogenesis studies of pentachlorophenol in F344/N rats. National Toxicology Program, U.S. Department of Health and Human Services, Research Triangle Park, NC. NTP TR 483, NIH No. 99-3973.

OEHHA (2006a). Child-specific reference doses (chRDs) for school site risk assessment, manganese and pentachlorophenol. Office of Environmental Health Hazard Assessment, California Environmental Protection Agency, Sacramento, CA, June 2006.

OEHHA (2006b). Proposition 65 Safe Harbor Levels: No significant risk levels for carcinogens and maximum allowable dose levels for chemicals causing reproductive toxicity. Office of Environmental Health Hazard Assessment, California Environmental Protection Agency, Oakland and Sacramento, August 2006.

Proudfoot A (2003). Pentachlorophenol poisoning. Toxicol Rev 22:3-11.

Sun H, Xu L, Chen J, Song L, Wang X (2006). Effect of bisphenol A, tetrachlorobisphenol A and pentachlorophenol on the transcriptional activities of androgen receptor-mediated reporter gene. Food Chem Toxicol 44:1916-1921.

Taylor T, Tucker T, Whalen M (2005). Persistent inhibition of human natural killer cell function by Ziram and pentachlorophenol. Environ Toxicol 20:418-424.

Terasaka S, Inoue A, Tanji M, Kiyama R (2006). Expression profiling of estrogen-responsive genes in breast cancer cells treated with alkylphenols, chlorinated phenols, parabens, or bis- and benzoylphenols for evaluation of estrogenic activity. Toxicol Lett 163:130-141.

U.S. EPA (2004). Reregistration Eligibility Decision for Pentachlorophenol. U.S. Environmental Protection Agency, Washington, DC. Docket ID No. EPA-HQ-OPP-2004-0402.

U.S. EPA (2007). Pentachlorophenol and its use as a wood preservative. U.S. Environmental Protection Agency, Washington, D.C. Accessed at: http://www.epa.gov/pesticides/factsheets/chemicals/pentachlorophenol main.htm.

Walls C, Glass W, Pearce N (1998). Health effects of occupational pentachlorophenol exposure in timber sawmill employees: a preliminary study. N Z Med J 111:362-364.

Zhao B, Yang J, Liu Z, Xu Z, Qiu Y, Sheng G (2006). Joint anti-estrogenic effects of PCP and TCDD in primary cultures of juvenile goldfish hepatocytes using vitellogenin as a biomarker. Chemosphere 65:359-364.

Picloram

A PHG of 500 ppb was developed for picloram (4-amino-3,5,6-trichloropicolinic acid) in drinking water in 1997. Picloram is a polychlorinated herbicide that has been widely used for the control of broad-leaved weeds and woody plants along rights-of-way. It is applied alone or in combination with 2,4-dichlorophenoxyacetic acid (2,4-D) in a product named Tordon.

Picloram is chemically related to the herbicides clopyralid and triclopyr; these chemicals are classified as auxins, which are plant growth regulators. Like other herbicides such as 2,4-D, these chemicals disrupt plant growth regulation, leading to overgrowth and death of susceptible plants. Mammalian toxicity is low. The PHG was based on increased liver weight in a 6-month dog-feeding study with a no-observed-adverse-effect level of 7 mg/kg-day and a lowest-observed-effect level of 35 mg/kg-day. Cancer studies have been negative except in some cases for liver tumors that were attributed to contamination of the administered product with hexachlorobenzene.

References

Blakley BR (1997). Effect of Roundup and Tordon 202C herbicides on antibody production in mice. Vet Hum Toxicol 39(4):204-6.

Jugulam M, McLean MD, Hall JC (2005). Inheritance of picloram and 2,4-D resistance in wild mustard (Brassica kaber). Weed Science 53:417–423.

Oakes DJ, Pollack JK (1999). Effects of a herbicide formulation, Tordon 75D, and its individual components on the oxidative functions of mitochondria. Toxicology 136(1):41-52.

Oakes DJ, Webster WS, Brown-Woodman PD, Ritchie HE (2002). A study of the potential for a herbicide formulation containing 2,4-D and picloram to cause malemediated developmental toxicity in rats. Toxicol Sci 68(1):200-6.

Oakes DJ, Webster WS, Brown-Woodman PD, Ritchie HE (2002). Testicular changes induced by chronic exposure to the herbicide formulation, Tordon 75D (2,4-dichlorophenoxyacetic acid and picloram) in rats. Reprod Toxicol 16(3):281-9.

U.S. EPA (1995). Reregistration Eligibility Decision (RED): Picloram. Prevention, Pesticides and Toxic Substances Office, U.S. Environmental Protection Agency, Washington, DC. August 1995. EPA 738-R95-019. Accessed at: http://www.epa.gov/REDs/0096.pdf.

1,2,4-Trichlorobenzene

A PHG of 0.005 mg/L (0.005 ppm, or 5 ppb) was developed in 1999 for 1,2,4-trichlorobenzene (1,2,4-TCB) in drinking water, based on enlargement of adrenal glands in a subchronic rat study. 1,2,4-TCB is used as a solvent in chemical manufacturing, a dye carrier for textiles, an intermediate in the production of other chemicals, a degreasing agent, a component of dielectric fluids, and as a component of lubricants and oils.

OEHHA recently reviewed 1,2,4-TCP for potential listing as a carcinogen under Proposition 65. A chronic study showed that male and female mice administered 1,2,4-TCB in their diet developed a high incidence of hepatocellular adenomas and carcinomas. A similar study in rats showed no evidence of carcinogenic effects, and a skin painting study in mice produced no evidence of carcinogenicity. Genotoxicity tests have mostly been negative, and dose-related increases in micronuclei have been negative. Considering this evidence, the expert committee concluded that 1,2,4-TCP should not be listed as a human carcinogen.

The 1999 PHG included a 10-fold factor for potential carcinogenicity of 1,2,4-TCB, based on preliminary reports of the 1994 CMA studies, which resulted in a total uncertainty factor (UF) of 10,000. Our present practice is to limit the total UF to 3,000. OEHHA acknowledges the potential concern about carcinogenicity from exposure to this halogenated benzene compound, and solicits further input on the approach for the planned PHG update.

References

Bogaards JJ, van Ommen B, Wolf CR, van Bladeren PJ (1995). Human cytochrome P450 enzyme selectivities in the oxidation of chlorinated benzenes. Toxicol Appl Pharmacol 132(1):44-52.

CMA (1994a). 104-Week Dietary Carcinogenicity Study with 1,2,4-Trichlorobenzene in Mice. Hazleton Washington, Inc., for the Chemical Manufacturers Association (Final Report, with cover letter dated 6/15/94).

CMA (1994b). 104-Week Dietary Carcinogenicity Study With 1,2,4-Trichlorobenzene in Rats. Hazleton Washington, Inc., for the Chemical Manufacturers Association (Final Report, with cover letter dated 6/15/94).

den Besten C, Smink MC, de Vries EJ, van Bladeren PJ (1991). Metabolic activation of 1,2,4-trichlorobenzene and pentachlorobenzene by rat liver microsomes: a major role for quinone metabolites. Toxicol Appl Pharmacol 108(2):223-33.

den Besten C, Vet JJ, Besselink HT, Kiel GS, van Berkel BJ, Beems R, van Bladeren PJ (1991). The liver, kidney, and thyroid toxicity of chlorinated benzenes. Toxicol Appl Pharmacol 111(1):69-81.

Kato Y, Kimura R (2002). The contribution of 2,3,5-trichlorophenyl methyl sulfone, a metabolite of 1,2,4-trichlorobenzene, to the delta-aminolevulinic acid synthetase induction by 1,2,4-trichlorobenzene in rat liver. Chemosphere 47(1):1-7.

Mizutani T, Miyamoto Y (1999). Modulation of halobenzene-induced hepatotoxicity by DT-diaphorase modulators, butylated hydroxyanisole and dicumarol: evidence for possible involvement of quinone metabolites in the toxicity of halobenzenes. Toxicol Lett 105:25-30.

OEHHA (2005). Evidence on the carcinogenicity of 1,2,4-trichlorobenzenze – Final, September, 2005. Reproductive and Cancer Hazard Assessment Branch, Office of Environmental Health Hazard Assessment, California Environmental Protection Agency, Oakland, CA. Accessed at:

www.oehha.ca.gov/prop65/hazard ident/pdf zip/HID124Trichlorobenzene final.pdf.

Trichlorofluoromethane (Freon 11)

A PHG of 0.7 mg/L (700 ppb) for trichlorofluoromethane (Freon 11) was developed in 1997. Freon 11 was one of the most widely used chlorofluorocarbons in industrial applications, including use as a blowing agent in foam production and as an aerosol propellant. Because of its effects on the atmospheric ozone layer, U.S. production was banned in 1996. A California PHG and MCL were developed because of its leaching from waste sites, and potential for water supply contamination.

The most well-known toxic effects of Freon 11 include cardiac and pulmonary disturbances (e.g., cardiac arrhythmias, tachycardia and hypotension) and changes in respiratory parameters, which were elucidated because of inhalation abuse of aerosol products. Other effects include hepatic lesions, central nervous system dysfunction, and skin and eye irritation. Chronic animal exposure studies were negative for carcinogenicity. Limited information is available on other potential effects, and our preliminary literature survey revealed no new toxicity studies.

References

Vinegar A (2001). PBPK modeling of canine inhalation exposures to halogenated hydrocarbons. Toxicol Sci 60(1):20-7.

Trichlorotrifluoroethane (Freon 113)

A Public Health Goal (PHG) of 4 mg/L (4 ppm) was developed for 1,1,2-trichloro-1,2,2-trifluoroethane (Freon 113) in drinking water in 1997. Freon 113 was widely used in industrial applications as a solvent for degreasing and dry cleaning, as a refrigerant, in

fire extinguishers, as a chemical intermediate and as a blowing agent in foam production. Because of its effects on the atmospheric ozone layer, U.S. production was banned in 1996. Exposures continued due to its persistence in air and presence in waste-water streams and leachate from waste sites, leading to the development of the California PHG and MCL.

Freon 113 has extremely low acute toxicity, but cardiac sensitization was reported from inhalation abuse of aerosol products, and confirmed in dog studies. Repeated short-term administration in several species did not reveal any distinctive toxicological effects. A rat chronic inhalation study resulted in increases in liver weight, but no carcinogenicity.

References

Bloom TF, Egeland GM (1991). Health-hazard evaluation report HETA 89-344-2157, Wiltech of Florida, Inc., Kennedy Space Center, Florida, Rothe Development, Inc., Johnson Space Center, Texas, report prepared for National Inst. for Occupational Safety and Health, Cincinnati, OH. OSTI Identifier OSTI ID: 5317162, Report No. PB-92-145895/XAB.

Eilers H, Kindler CH, Bickler PE (1999). Different effects of volatile anesthetics and polyhalogenated alkanes on depolarization-evoked glutamate release in rat cortical brain slices. Anesth Analg 88(5):1168-74.

Neghab M, Qu S, Bai CL, Caples J, Stacey NH (1997). Raised concentration of serum bile acids following occupational exposure to halogenated solvents, 1,1,2-trichloro-1,2,2-trifluoroethane and trichloroethylene. Int Arch Occup Environ Health 70(3):187-94.